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# Signal Transduction by Neurotransmitters in Brain and Heart in Health and Disease

### Position

Professor of Physiology, Sackler Faculty of Medicine

### Research

Electrical activity of excitable cells is their most important feature, which allows the performance of fundamental functions of brain, heart and muscle. We are addressing a key issue in modern cardiology and neurobiology: how neurotransmitters regulate cardiac cells and neurons by acting on ion channels – proteins that underlie the electrical activity in these cells; and how errors in these processes cause disease. Main projects in the lab:

Function and regulation of receptors, G proteins, Ca<sup>2+</sup> and K<sup>+</sup> channels in health and disease; Ion channelrelated hereditary cardiac and neurological disorders (channelopathies); Mechanisms of coupling of G protein-coupled receptors with effectors; Molecular mechanisms of bipolar disorder.



**Research methods:** Electrophysiology, Neurophysiology, Heterologous Expression, Protein Biochemistry, Fluorescence Resonance Energy Transfer (FRET), Molecular biology, Mathematical and Kinetic Modeling and Simulation, Immunocytochemistry

### Publications

Babai N, Kanevsky N, **Dascal N**, Rozanski GJ, Singh DP, Fatma N & Thoreson WB (2010). Anion sensitive regions of L-type  $Ca_v 1.2$  calcium channels expressed in HEK293 cells. *PLoS One*, 5, e8602.

Berlin S, Keren-Raifman T, Castel R, Rubinstein M, Dessauer CW, Ivanina T & **Dascal N** (2010).  $G\alpha_i$ and  $G\beta\gamma$  jointly regulate the conformations of a  $G\beta\gamma$ effector, the neuronal G-protein activated K<sup>+</sup> channel (GIRK). *J Biol Chem*, 285, 6179-6185.



Studying GIRK channels expressed in a heterologous system (Xenopus oocytes). Intramolecular fluorescence resonance energy transfer (i-FRET) shows interactions of cytosolic N- and C-termini of the channel. **A**, GIRK channel labeled with two fluorescent proteins. **B**, Imaging the expressed fluorescent proteins with a confocal microscope. **C**, **D**, Example of use of FRET analysis to study conformational changes in the channel caused by neurotransmitter, G proteins or drugs. **E**, G $\alpha$  and G $\beta\gamma$  synergistically alter the conformation of GIRK1 subunit. Edelheit O, Hanukoglu I, Shriki Y, Tfilin M, **Dascal N**, Gillis D & Hanukoglu A (2010). Truncated  $\beta$  epithelial sodium channel (ENaC) subunits responsible for multi-system pseudohypoaldosteronism support partial activity of ENaC. *J Steroid Biochem Mol Biol*, 119, 84-88.

Tselniker I, Tsemakhovich VA, Dessauer CW & **Dascal N**. (2010) Stargazin modulates neuronal voltagedependent Ca<sup>2+</sup> channel Ca<sub>v</sub>2.2 by a G $\beta\gamma$ -dependent mechanism. *J Biol Chem* **285**, 20462-20471.

Laish-Farkash A, Brass D, Marek-Yagel D, Pras E, **Dascal N**, Antzelevitch C, Nof E, Reznik H, Glikson M & Luria D (2010). A novel mutation in the HCN4 gene causes symptomatic sinus bradycardia in Moroccan Jews. *J Cardiovasc Electrophysiol* 21, 1365-1372.

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Oz S, Tsemakhovich V, Christel CJ, Lee A & **Dascal** N. (2011). CaBP1 regulates voltage dependent inactivation and activation of  $Ca_v 1.2$  (L-type) calcium channels. *J Biol Chem* 286, 13945-13953.

Edelheit O, Hanukoglu I, **Dascal N** & Hanukoglu A. (2011) Identification of the roles of conserved charged residues in the extracellular domain of an epithelial sodium channel (ENaC) subunit by alanine mutagenesis. *Am J Physiol Renal Physiol* 300, F887-897.

Berlin S, Tsemakhovich VA, Castel R, Ivanina T, Dessauer CW, Keren-Raifman T & **Dascal N**. (2011) Two distinct aspects of coupling between  $G\alpha_i$  and G protein-activated K<sup>+</sup> channel (GIRK) revealed by fluorescently-labeled  $G\alpha_{i3}$  subunits. *J Biol Chem* 287, 19537-19549.

Almagor L, Chomsky-Hecht O, Ben-Mocha A, Hendin-Barak D, **Dascal N** & Hirsch JA. (2012). The role of a voltage-dependent Ca<sup>2+</sup> channel intracellular linker: a structure-function analysis. *J Neurosci* 32, 7602-7613.

Pankonien I, Otto A, **Dascal N**, Morano I & Haase H. (2012). Ahnak1 interaction is affected by phosphorylation of Ser-296 on  $Ca_{\nu\beta}2$ . *Biochem Biophys Res Commun* 421, 184-189.

Weiss S, Keren-Raifman T, Oz S, Ben Mocha A, Haase H & **Dascal N**. (2012). Modulation of distinct isoforms of L-type calcium channels by  $G_q$ -coupled receptors in *Xenopus* oocytes: Antagonistic effects of G $\beta\gamma$  and protein kinase C. *Channels* **6**, 426-437.

Almagor L, Chomsky-Hecht O, Ben Mocha A, Hendin-Barak D, **Dascal N** & Hirsch JA. (2012). Ca<sub>v</sub>1.2 I-II linker structure and Timothy syndrome. *Channels* **6**, 468-472.

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Weisbrod, D., Peretz, A., Ziskind, A., Menaker, N., Oz, S., Barad, L., Eliyahu, S., Itskovitz-Eldor, J., **Dascal, N.**, Khananshvili, D., Binah, O., and Attali, B. (2013) SK4 Ca<sup>2+</sup> activated K<sup>+</sup> channel is a critical player in cardiac pacemaker derived from human embryonic stem cells. *Proc Natl Acad Sci USA* 110, E1685-1694.

Weiss S, Oz S, Benmocha A, **Dascal N**. (2013) Regulation of cardiac L-type Ca<sup>2+</sup> channel Ca<sub>v</sub>1.2 via the  $\beta$ -adrenergic-cAMP-protein kinase A pathway: old dogmas, advances, and new uncertainties. *Circ Res* 2013, 113:617-31.

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Edelheit O, Ben-Shahar R, **Dascal N**, Hanukoglu A & Hanukoglu I. (2014). Conserved charged residues at the surface and interface of epithelial sodium channel (ENaC) subunits: roles in cell surface expression and Na<sup>+</sup> self-inhibition response. *FEBS J*. 281:2097-111.

## Grants

2013-2016 Mechanisms of isoform-specific regulation of L-type Ca<sup>2+</sup> channels by protein kinases. German-Israel Foundation (GIF), With S. Weiss and E. Klussmann.