Bone-to-Implant Contact and New Bone Formation Within Human Freeze-Dried Bone Blocks Grafted Over Rabbit Calvaria

Zvi Artzi, DMD¹/Karen Anavi-Lev, DMD²/Avital Kozlovsky, DMD³/ Liat Chaushu, DMD⁴/Frank Schwarz, DMD⁵/Haim Tal, DMD, PhD⁶

Purpose: To assess the extent of osseointegration with rough-surface implants and new bone formation (NBF) within human freeze-dried bone blocks (h-FDB) grafted over rabbit calvaria. Materials and Methods: A total of 18 rectangular h-FDB blocks were stabilized bilaterally to the calvaria of nine New Zealand rabbits by two mini titanium screws each. A total of 18 rough-surface implants (5.0×6.0 -mm) were placed, 9 simultaneously (immediate placement [IP]) on one side and 9 at 3 months after block grafting (delayed placement [DP]) on the contralateral side. At 12 weeks after the second surgical procedure, block biopsies were harvested and processed for histologic analysis. Morphometric measurements consisted of bone-to-implant contact (BIC) and the extent of NBF from the calvarial surface and outward into the block. A paired t test was applied for statistical analysis. Results: All h-FDB blocks were integrated, and the implants showed clinical stability. Histologically, the BIC was primarily between the apical end of the implants and the host rabbit calvaria. Bone growth between the implant threads was minimal and inconsistent among all animals. Morphometric measurements showed that the mean BIC of the IP and DP implants with the blocks was 10.50% \pm 5.99% and 23.06% \pm 9.58%, respectively (P < .001). NBF was observed primarily in the cancellous compartment of the block adjacent to the recipient calvarial bed. The extent of NBF into the block around the IP and DP implants was 9.95% \pm 8.41% and 12.90% \pm 11.07%, respectively (P = 0.2). Conclusion: In this model, a significantly lower BIC was demonstrated when implants were placed simultaneously with h-FDB block grafting compared to those placed in a two-stage mode. However, both techniques showed limited osseointegration. INT J ORAL MAXILLOFAC IMPLANTS 2017;32:768-773. doi: 10.11607/jomi.5366

Keywords: bone-implant contact, FDB bone blocks, freeze-dried bone, graft vascularization, new bone formation, osseointegration

¹Professor of Periodontology; Director of Graduate Periodontics, Department of Periodontology and Oral Implantology, School of Dental Medicine, Tel Aviv University, Tel Aviv, Israel.

²Periodontist, Master (Msc) in Periodontology, Department of Periodontology and Oral Implantology, School of Dental Medicine, Tel Aviv University, Tel Aviv, Israel.

- ³Associate Professor, Department Coordinator, Department of Periodontology and Oral Implantology, School of Dental Medicine, Tel Aviv University, Tel Aviv, Israel.
- ⁴Post-Graduate Student in Periodontology, Department Coordinator, Department of Periodontology and Oral Implantology, School of Dental Medicine, Tel Aviv University, Tel Aviv, Israel.
- ⁵Professor of Oral Medicine and Peri-implant Infections, Department of Periodontology and Oral Implantology, School of Dental Medicine, Tel Aviv University, Tel Aviv, Israel.
- ⁶Professor of Periodontology, Chairman of the Department of Periodontology and Oral Implants, Department of Oral Surgery, Heinrich Heine University, Düsseldorf, Germany.

Avital Kozlovsky and Haim Tal equally contributed to this study.

Correspondence to: Dr Zvi Artzi, Department of Periodontology and Oral Implantology, School of Dental Medicine, Tel Aviv University, Tel Aviv, Israel 69978. Fax: +972-35402768. Email: zviartzi@tau.ac.il

©2017 by Quintessence Publishing Co Inc.

lveolar bone deficiency is one of the major ob-Astacles in implant reconstructions, often requiring restoration of the alveolar process via bone augmentation procedures. The application of autogenous bone block transplantation has a predictable outcome and well-established long-term success¹⁻⁵; however, graft resorption,⁶⁻⁹ donor site limitations, and morbidity considerations^{8,10–14} call for alternative approaches. Consequently, other sources of allogeneic and xenogeneic origins have been proposed.^{15–29} Clinical reports on the use of onlay human freeze-dried bone (h-FDB) blocks with subsequent implant placement have shown promising clinical results.^{15,30–36} However, no controlled trials, comparative studies, or histologic analyses have been conducted to validate the efficacy of these grafted biomaterials.

In their systematic review,³⁷ Waasdorp and Reynolds gathered literature on h-FDB onlay grafts for alveolar ridge augmentation from 1950 to 2008. Only nine publications met their inclusion criteria: two case



Fig 1 Two h-FDB blocks were stabilized by fixation screws followed by an immediately placed (IP) implant placement at one of the grafted blocks.

reports and seven case series with short-term followup. No randomized controlled clinical trials (RCTs) were identified in the search. Those authors concluded that there is insufficient evidence to establish treatment efficacy of graft incorporation, alveolar ridge augmentation, and long-term dental implant survival.

A recent systematic review of the literature³⁸ provided updated histomorphometric and histologic characteristics of h-FDB blocks. Of the 15 articles that met the inclusion criteria (361 blocks), there was not a single report on the quality/quantity of the osseointegration. Only two studies^{27,36} that evaluated fresh-frozen osseous blocks also included a control group, and both reported that the majority of slides demonstrated large numbers of empty osteocyte lacunae in non-vital segments of necrotic bone and no direct contact between remodeled and grafted bone.

In the present study, h-FDB blocks were used as onlay grafts over rabbit calvaria. The rabbit skull has been used extensively as an appropriate site to observe and analyze grafted biomaterials,^{39–44} including xenografts, and animal models are well accepted for the analyses of these blocks both histologically and morphometrically.⁴² Minimal morbidity of the animal, ease of access to the site, and predictability of soft tissue management over the augmented site are just a few advantages of choosing this animal model. Therefore, the aim of this study was to evaluate the amount of direct bone-toimplant contact (BIC) between the h-FDB blocks and the implants, which were placed either simultaneously or in a delayed two-stage fashion. The different placement methods were to allow testing of whether the timing of implant placement has any impact in a non-integrated block vs a 3-month integrated block. The extent of new bone formation (NBF) in these blocks and the nature of the osseous connection between the grafted blocks and the recipient calvarial beds were also assessed.



Fig 2 An additional implant was placed at the second block on the contralateral side at 3 months following the h-FDB grafting phase.

MATERIALS AND METHODS

The institutional committee of animal care of Tel Aviv University approved the study. The study comprised 10 New Zealand female rabbits aged from 4 to 6 months weighing from 2.5 to 3.0 kg. They were kept in a calm secluded room in separate cages, fed Teklad Global Rabbit Diet (Envigo) daily, and given tap water ad libitum.

The surgical procedures were performed under general anesthesia following pre-sedation with 1.5 cc (20 mg) 2% xylazine base IM (Sedaxylan Veterinary, Eurovet Animal Health BV), followed by an IV combination of ketamine (Clorketam, Vetoquinol) 5 mg/kg + xylazine base (XYL – M 2, Veterinary) 1 mg/kg. In addition, a transdermal slow release (50 µg/h) sticky patch of fentanyl 8.25 mg (Novosis AG) was adhered to the rabbit's shaved upper back for 3 days.

Local infiltration of 2% lidocaine hydrochloride with norepinephrine (1:100,000) was administered for hemostasis and reduction of postoperative pain.

Once anesthetized, the rabbit calvarium was exposed via a midsagittal longitudinal incision. Full-thickness dermal flaps were reflected, exposing the calvarial cortex. Cortical perforations were established by a 1-mm rounded diamond burr to increase vascular flow at the relevant site. In each calvarium, a pair of 10 mm (W) \times 10 mm (L) \times 5 mm (H) rectangular corticocancellous h-FDB blocks (LifeNet Health, Inc) were adjusted and placed on the rabbit's parietal calvarial surface bilaterally. Blocks were stabilized by two 1.2-mm titanium screws (Osteomed) in order to establish an intimate contact with the external calvarial surface. Next a randomly allocated (via coin flip) implant site was prepared and an implant (5.0 mm \times 6.0 mm; ATID, Alpha-Bio Tec Ltd) placed. This was referred to as the immediately placed (IP) implant (Fig 1).

The surgical wounds were closed by suturing in layers. The periosteal margins were approximated by



Fig 3 A nondecalcification section of an IP implant in the h-FDB. The BIC is evident primarily at the calvarial host bed (H) and to a lesser degree at the cancellous portion of the block (BL) on one side only. There was no observable contact at the cortical layer zone of the block (nondecalcified toluidine blue staining, $\times 17.5$ magnification).

simple interrupted suture after releasing the flaps by periosteal incisions parallel to the main surgical one. A primary non-tensional soft-tissue closure was obtained using interrupted horizontal internal mattress suture followed by a continuous interlock suture. All sutures were made using a 5-0 resorbable Vicryl suture. At 3 months following this first phase of the surgery, a second implant, which was referred to as the delayed placed (DP) implant, was placed on the contralateral block (Fig 2).

An effort was made to place the implants at the block level while 1 mm of the most apical portion of the implant would be placed in the native calvarial bone. However, with this method, the implant platform might be left slightly higher than the level of the block.

Postoperative antibiotics were given for 3 consecutive days after each surgical intervention (0.5 cc of durabiotic 5% IM, Baytril, Bayer AG). During follow-up, one rabbit showed signs of distress and started to lose weight. Consequently, this animal was euthanized and dropped from the study, leaving nine rabbits available for study.

At 6 months following the first surgical procedure, the animals were injected with ketamine (1 cc) + xylazine (1.5 cc) followed by a lethal dose (30 mg/kg) of pentobarbitone sodium 200 mg/ml IV (CTS, Pharmaceutical Industries Inc). The animals were decapitated, and the surgical sites were retrieved en bloc. The specimens were put in 10% buffered formalin for 1 week and then transferred to a 70% ethanol solution. Radiographs of the specimen blocks were taken before further histologic processing.

Histologic Processing

Nine calvaria, providing 18 specimens total, were available for non-decalcified histologic processing,⁴⁵ which was performed according to a



Fig 4 A nondecalcification section of a DP implant in the h-FDB. The BIC is clearly observed inside the threads of the implant at the cancellous portion of the block (BL), however, there is no osseointegration at the cortical zone of the block (nondecalcified toluidine blue staining, \times 17.5 magnification). H = calvarial host bed.

standardized procedure.⁴⁶ In brief, tissue biopsies were dehydrated using ascending grades of alcohol and xylene, infiltrated, and embedded in methylmethacrylate (Technovit 9100 NEU, Heraeus Kulzer). Three sections approximately 300 μ m in thickness were obtained per block at the most central aspect of the titanium implant, and both osteosynthesis screws were fixed in place using a diamond band saw (Exakt, Apparatebau). The sections were ground to a final thickness of approximately 40 μ m and stained with toluidine blue.

All measurements were jointly taken by two investigators (Z.A. and K.A.L.) while the identification of the site was masked. Histomorphometry was conducted on a screen monitor attached to the microscope (magnification \times 35) using the Bioquant Nova Prime System (Bioquant Image Analysis Corp) software.

Direct BIC was compared between the IP and DP implants within the surrounding new bone out of the total block housing and particularly at its cancellous area (or portion). The BIC was also measured at the calvarial implant zone to be used as a reference. New bone formation (NBF) was calculated as the percentage of distance penetration of newly formed osseous tissue out of the total vertical dimension of the h-FDB. This was achieved by the mean measurements taken from peripheral mineralized stained areas proximal to the implants and proximal to the fixation screws close to the observed BIC. Also, a demarcation between the outer calvarial bony envelope and pale staining of osteoid formation served as accessory tools to distinguish between the host and the block mineralized zones. Apart from the IP and DP implants, the fixation screws, which represent machined-surface titanium, were also evaluated and recorded.

Table 1Morphologic Assessment Comparing New Bone Formation (NBF) and Bone-to-ImplantContact (BIC) at Immediate (IP) and Delayed (DP) Placed Implants and at theFixation Screws					
Implant/screw	BIC/calvaria (%)	BIC/blocks (%)	BIC/cancellous (%)	NBF/blocks (%)	NBF/cancellous (%)
IP implants $(n = 9)$	70.7	10.5	14.6	9.9	15.3
DP implants $(n = 9)$	72.9	23.1	35.7	12.9	19.7
Fixation screws (n = 35)	79.6	10.8	15.0	N/A	N/A

Calvaria = the host bed; block = trabecular + corticalis; cancellous = only the trabecular zone; N/A = not applicable.

Statistical analysis was performed using the paired *t* test.

RESULTS

Histology and Histomorphometry

NBF was observed primarily in the cancellous compartment of the block adjacent to the recipient calvarial bed. Bone growth between the implant threads was limited and inconsistent in all specimens. The BIC was observed primarily between the apical end of the implant and the calvarium. The BIC within the h-FDB block was established mainly at the implant threads proximal to the host bone. Neither BIC nor bone growth into the threads was evident at the cortical zone of the block. The graft matrix showed no signs of osteoclastic or other resorption, and the osteocyte lacunae were, in general, empty (Figs 3 and 4).

The BIC in the IP and DP implant groups was 10.50% \pm 5.99% and 23.06% \pm 9.58%, respectively (Table 1). The differences were statistically significant (*P* < .001). Excluding the cortical portion, the BIC at the cancellous portion was 14.61% \pm 7.51% and 35.67% \pm 16.14% for the IP and DP implants, respectively (*P* < .002).

The BIC at the calvarial zone was significantly higher than the BIC within the block, with the mean grade varying from 68.11% \pm 12.00% for the IP implants to 74.73% \pm 19.36% for the DP implants. However, there was no significant difference between the groups. The NBF, indicating the vertical extent of bone formation into the block scaffold from the calvarial surface, was 9.95% \pm 8.41% and 12.90% \pm 11.07%, respectively (P = .2). After excluding the cortical non-vital zone of the blocks, the NBF was 15.30% \pm 13.81% and 19.71% \pm 16.59%, respectively (P = .216). There was no significant correlation between the BIC and the NBF in any of the groups.

It is noteworthy that the BIC between the machined surface titanium fixation screws and the bone growth was $10.8\% \pm 5.23\%$ within the h-FDB and $15.0\% \pm 16.18\%$ at its cancellous portion (Fig 5). The BIC to the fixation screws at the calvarial zone was remarkably higher (79.6% \pm 27.3%; P < .0001).



Fig 5 A nondecalcification section of the titanium fixation screw in the h-FDB. The BIC is evident only at the calvarial host bed (H) and in the proximal area of the cancellous portion (Can) of the block. There is no sign of osseointegration at the cortical layer (Cor) of the block (nondecalcified toluidine blue staining, \times 17.5 magnification).

DISCUSSION

This study was undertaken to investigate the potential use of h-FDB onlay grafts for alveolar ridge augmentation. Block stabilization was achieved by two fixation screws to a cortical perforated intramembranous osseous bed, which allowed vascular passage into the cancellous part of the block. Implants were placed either simultaneously (IP) or after 3 months of block integration (DP); thus, the establishment of osseointegration was assessed in two different non-vital/viable tissue surroundings. Clinically, all h-FDB blocks had been well integrated. At 6 months, the NBF as well as some sparse BIC around the implants' titanium surfaces were evident. Histologically, the amount of NBF in the blocks was quite limited. Both NBF and BIC were observed primarily near the calvarial bed. Since there was only an average of 10% to 13% of NBF ingrowth in the h-FDB, most of the block became a non-vital scaffold. The middle and coronal parts of the implant's osseous housing remained non-vital, and no osseointegration process could be identified. A similar outcome was recently reported in humans who received fresh-frozen allogeneic bone.^{28,29}

An appropriate biomaterial scaffold should allow vascular development. The fact that there is no evidence of vascularization in the block margins (ie, the compact cortex) would clinically eventually prevent vitalization, followed by inability to establish NBF in this particular area. Apparently, the perfusion of vessels within this region is obstructed. This might explain in part the difference between the cortical and the trabecular zones in terms of tissue replacement. Currently, the present authors are investigating tracing the vascular formation rate and angiogenesis in these grafted blocks in a similar animal model by tetracycline and calcein labeling.

Implants were placed in their apical zones at the native/host bone. This was to assure initial stability and to initiate an immediate osseointegration process.

In addition, the calvarial cortical perforations enabled enhancement of vascular pathway to the stabilized attached block.

Implants were placed at two points in time: simultaneously with the blocks and at 3 months following block grafting. The outcomes of both BIC and NBF showed that the two-stage approach had an advantage over the combined technique—there was a significant difference in favor of the DP implants over the IP implants. This could be attributed to the fact that the DP implants were installed in a 3-month partly vital h-FDB, which provides better vascularization and better mechanical stability at the portion in which there is direct contact between the pristine calvarial bone and the implant surface.

In general, the greater BIC over NBF could be related to the fact that the rough surface of the implant serves as an osteoconductive vehicle, which is not proven as related to the blocks themselves. Apparently, the combination of placing an implant surface that has been proven by evidence to be osteoconductive in an already remodeled and viable grafted block would be the timing of the ideal treatment approach.

Previous studies with autogenous blocks^{47,48} and/or particulate biomaterials⁴⁹ have also shown a discernible difference in the amount of BIC and NBF in favor of the DP approach. It would appear that augmented bone could serve as appropriate osseous housing for an osseointegrated implant, provided that the grafted biomaterial (h-FDB) had first been revascularized and repopulated by new bone growth, thus making it comparable to a pristine alveolar ridge.

The lack of long-term evidence-based data, as reflected in current reviews,^{50,51} warrants in-depth methodical future research to determine the long-term efficacy of h-FDB in terms of stable osseointegration and the capability of total new bone growth replacement.

CONCLUSIONS

The experimental animal model used in the current study demonstrated limited and inconsistent new bone growth into human freeze-dried corticocancellous bone blocks. This indicates that success of osseointegration of implants placed in these block grafts in augmentation procedures is probably uncertain.

ACKNOWLEDGMENTS

The study was supported by Turnheim Research Funds. The authors thank Ms Tina Hagena for the histological processing, Mr Haim Berky for the animal care, Ms Ilana Gelernter for the statistical analysis, and Ms Esther Eshkol for the manuscript editing. The authors declare that they have no conflicts of interest.

REFERENCES

- Adell R, Lekholm U, Gröndahl K, Brånemark PI, Lindström J, Jacobsson M. Reconstruction of severely resorbed edentulous maxillae using osseointegrated fixtures in immediate autogenous bone grafts. Int J Oral Maxillofac Implants 1990;5:233–246.
- Isaksson S, Alberius P. Maxillary alveolar ridge augmentation with onlay bone-grafts and immediate endosseous implants. J Craniomaxillofac Surg 1992;20:2–7.
- Astrand P, Nord PG, Brånemark PI. Titanium implants and onlay bone graft to the atrophic edentulous maxilla: A 3-year longitudinal study. Int J Oral Maxillofac Surg 1996;25:25–29.
- 4. van Steenberghe D, Naert I, Bossuyt M, et al. The rehabilitation of the severely resorbed maxilla by simultaneous placement of autogenous bone grafts and implants: A 10-year evaluation. Clin Oral Investig 1997;1:102–108.
- Keller EE, Tolman DE, Eckert S. Surgical-prosthodontic reconstruction of advanced maxillary bone compromise with autogenous onlay block bone grafts and osseointegrated endosseous implants: A 12-year study of 32 consecutive patients. Int J Oral Maxillofac Implants 1999;14:197–209.
- Davis WH, Martinoff JT, Kaminishi RM. Long-term follow up of transoral rib grafts for mandibular atrophy. J Oral Maxillofac Surg 1984;42:606–609.
- Widmark G, Andersson B, Ivanoff CJ. Mandibular bone graft in the anterior maxilla for single-tooth implants. Presentation of surgical method. Int J Oral Maxillofac Surg 1997;26:106–109.
- Cordaro L, Amadé DS, Cordaro M. Clinical results of alveolar ridge augmentation with mandibular block bone grafts in partially edentulous patients prior to implant placement. Clin Oral Implants Res 2002;13:103–111.
- Araújo MG, Sonohara M, Hayacibara R, Cardaropoli G, Lindhe J. Lateral ridge augmentation by the use of grafts comprised of autologous bone or a biomaterial. An experiment in the dog. J Clin Periodontol 2002;29:1122–1131.
- 10. Raghoebar GM, Louwerse C, Kalk WW, Vissink A. Morbidity of chin bone harvesting. Clin Oral Implants Res 2001;12:503–507.
- 11. Nkenke E, Schultze-Mosgau S, Radespiel-Tröger M, Kloss F, Neukam FW. Morbidity of harvesting of chin grafts: A prospective study. Clin Oral Implants Res 2001;12:495–502.

- Nkenke E, Weisbach V, Winckler E, et al. Morbidity of harvesting of bone grafts from the iliac crest for preprosthetic augmentation procedures: A prospective study. Int J Oral Maxillofac Surg 2004;33:157–163.
- Clavero J, Lundgren S. Ramus or chin grafts for maxillary sinus inlay and local onlay augmentation: Comparison of donor site morbidity and complications. Clin Implant Dent Relat Res 2003;5:154–160.
- von Arx T, Häfliger J, Chappuis V. Neurosensory disturbances following bone harvesting in the symphysis: A prospective clinical study. Clin Oral Implants Res 2005;16:432–439.
- Köndell PA, Mattsson T, Astrand P. Immunological responses to maxillary on-lay allogeneic bone grafts. Clin Oral Implants Res 1996;7:373–377.
- Lyford RH, Mills MP, Knapp CI, Scheyer ET, Mellonig JT. Clinical evaluation of freeze-dried block allografts for alveolar ridge augmentation: A case series. Int J Periodontics Restorative Dent 2003;23:417–425.
- 17. Leonetti JA, Koup R. Localized maxillary ridge augmentation with a block allograft for dental implant placement: Case reports. Implant Dent 2003;12:217–226.
- Keith JD Jr. Localized ridge augmentation with a block allograft followed by secondary implant placement: A case report. Int J Periodontics Restorative Dent 2004;24:11–17.
- Keith JD Jr, Petrungaro P, Leonetti JA, et al. Clinical and histologic evaluation of a mineralized block allograft: Results from the developmental period (2001-2004). Int J Periodontics Restorative Dent 2006;26:321–327.
- Barone A, Varanini P, Orlando B, Tonelli P, Covani U. Deep-frozen allogeneic onlay bone grafts for reconstruction of atrophic maxillary alveolar ridges: A preliminary study. J Oral Maxillofac Surg 2009;67:1300–1306.
- Rothamel D, Schwarz F, Herten M, et al. Vertical ridge augmentation using xenogenous bone blocks: A histomorphometric study in dogs. Int J Oral Maxillofac Implants 2009;24:243–250.
- Peleg M, Sawatari Y, Marx RN, et al. Use of corticocancellous allogeneic bone blocks for augmentation of alveolar bone defects. Int J Oral Maxillofac Implants 2010;25:153–162.
- 23. Schlee M, Rothamel D. Ridge augmentation using customized allogenic bone blocks: Proof of concept and histological findings. Implant Dent 2013;22:212–218.
- Nissan J, Marilena V, Gross O, Mardinger O, Chaushu G. Histomorphometric analysis following augmentation of the posterior mandible using cancellous bone-block allograft. J Biomed Mater Res A 2011;97:509–513.
- Nissan J, Mardinger O, Calderon S, Romanos GE, Chaushu G. Cancellous bone block allografts for the augmentation of the anterior atrophic maxilla. Clin Implant Dent Relat Res 2011;13:104–111.
- Nissan J, Marilena V, Gross O, Mardinger O, Chaushu G. Histomorphometric analysis following augmentation of the anterior atrophic maxilla with cancellous bone block allograft. Int J Oral Maxillofac Implants 2012;27:84–89.
- 27. Spin-Neto R, Landazuri Del Barrio RA, Pereira LA, Marcantonio RA, Marcantonio E, Marcantonio E Jr. Clinical similarities and histological diversity comparing fresh frozen onlay bone blocks allografts and autografts in human maxillary reconstruction. Clin Implant Dent Relat Res 2013;15:490–497.
- 28. Spin-Neto R, Stavropoulos A, Coletti FL, Faeda RS, Pereira LA, Marcantonio E Jr. Graft incorporation and implant osseointegration following the use of autologous and fresh-frozen allogeneic block bone grafts for lateral ridge augmentation. Clin Oral Implants Res 2014;25:226–233.
- Spin-Neto R, Stavropoulos A, Coletti FL, Pereira LA, Marcantonio E Jr, Wenzel A. Remodeling of cortical and corticocancellous freshfrozen allogeneic block bone grafts--a radiographic and histomorphometric comparison to autologous bone grafts. Clin Oral Implants Res 2015;26:747–752.
- Petrungaro PS, Amar S. Localized ridge augmentation with allogenic block grafts prior to implant placement: Case reports and histologic evaluations. Implant Dent 2005;14:139–148.
- Pendarvis WT, Sandifer JB. Localized ridge augmentation using a block allograft with subsequent implant placement: A case series. Int J Periodontics Restorative Dent 2008;28:509–515.

- 32. Chaushu G, Mardinger O, Calderon S, Moses O, Nissan J. The use of cancellous block allograft for sinus floor augmentation with simultaneous implant placement in the posterior atrophic maxilla. J Periodontol 2009;80:422–428.
- Chaushu G, Vered M, Mardinger O, Nissan J. Histomorphometric analysis after maxillary sinus floor augmentation using cancellous bone-block allograft. J Periodontol 2010;81:1147–1152.
- Contar CM, Sarot JR, Bordini J Jr, Galvão GH, Nicolau GV, Machado MA. Maxillary ridge augmentation with fresh-frozen bone allografts. J Oral Maxillofac Surg 2009;67:1280–1285.
- Contar CM, Sarot JR, da Costa MB, et al. Fresh-frozen bone allografts in maxillary ridge augmentation: Histologic analysis. J Oral Implantol 2011;37:223–231.
- Acocella A, Bertolai R, Ellis E 3rd, Nissan J, Sacco R. Maxillary alveolar ridge reconstruction with monocortical fresh-frozen bone blocks: A clinical, histological and histomorphometric study. J Craniomaxillofac Surg 2012;40:525–533.
- Waasdorp J, Reynolds MA. Allogeneic bone onlay grafts for alveolar ridge augmentation: A systematic review. Int J Oral Maxillofac Implants 2010;25:525–531.
- Monje A, Pikos MA, Chan HL, et al. On the feasibility of utilizing allogeneic bone blocks for atrophic maxillary augmentation. Biomed Res Int 2014;2014:814578.
- 39. Lundgren D, Lundgren AK, Sennerby L, Nyman S. Augmentation of intramembraneous bone beyond the skeletal envelope using an occlusive titanium barrier. An experimental study in the rabbit. Clin Oral Implants Res 1995;6:67–72.
- Mohammadi S, Rasmusson L, Göransson L, Sennerby L, Thomsen P, Kahnberg KE. Healing of titanium implants in onlay bone grafts: An experimental rabbit model. J Mater Sci Mater Med 2000;11:83–89.
- Tamini F, Torres J, Gbureck U, et al. Craniofacial vertical bone augmentation: A comparison between 3D printed monolithic monetite blocks and autologous onlay grafts in the rabbit. Biomaterials 2009;30:6318–6326.
- Kim SJ, Shin HS, Shin SW. Effect of bone block graft with rhBMP-2 on vertical bone augmentation. Int J Oral Maxillofac Surg 2010;39:883–888.
- 43. Torres J, Tamimi F, Alkhraisat MH, et al. Vertical bone augmentation with 3D-synthetic monetite blocks in the rabbit calvaria. J Clin Periodontol 2011;38:1147–1153.
- 44. Minami M, Takechi M, Ohta K, et al. Bone formation and osseointegration with titanium implant using granular- and blocktype porous hydroxyapatite ceramics (IP-CHA). Dent Mater J 2013;32:753–760.
- 45. Donath K, Breuner G. A method for the study of undecalcified bones and teeth with attached soft tissues. The Säge-Schliff (sawing and grinding) technique. J Oral Pathol 1982;11:318–326.
- 46. Schwarz F, Jung RE, Fienitz T, Wieland M, Becker J, Sager M. Impact of guided bone regeneration and defect dimension on wound healing at chemically modified hydrophilic titanium implant surfaces: An experimental study in dogs. J Clin Periodontol 2010;37:474–485.
- Lundgren S, Rasmusson L, Sjöström N, Sennerby L. Simultaneous or delayed placement of titanium implants in free autogenous iliac bone grafts. Histological analysis of the bone graft-titanium interface in 10 consecutive patients. Int J Oral Maxillofac Surg 1999;28:31–37.
- Rasmusson L, Meredith N, Kahnberg KE, Sennerby L. Stability assessments and histology of titanium implants placed simultaneously with autogenous onlay bone in the rabbit tibia. Int J Oral Maxillofac Surg 1998;27:229–235.
- 49. Artzi Z, Nemcovsky CE, Tal H, et al. Simultaneous versus twostage implant placement and guided bone regeneration in the canine: Histomorphometry at 8 and 16 months. J Clin Periodontol 2010;37:1029–1038.
- Fretwurst T, Gad LM, Nelson K, Schmelzeisen R. Dentoalveolar reconstruction: Modern approaches. Curr Opin Otolaryngol Head Neck Surg 2015;23:316–322.
- Leong DJ, Oh TJ, Benavides E, Al-Hezaimi K, Misch CE, Wang HL. Comparison between sandwich bone augmentation and allogenic block graft for vertical ridge augmentation in the posterior mandible. Implant Dent 2015;24:4–12.